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METHODS FOR INCREASING THE RATE OF HEART MUSCLE CONTRACTION

Cross Reference

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This application claims priority to U.S. Provisional Patent Application Serial Nos. 60/405,199 filed August 22, 2002 and 60/448,953 filed February 21, 2003, both of which are incorporated by reference herein in their entirety.

Field of Invention

This invention relates generally to methods for increasing the contractile rate in heart muscle and methods for treating and preventing cardiac disorders.

10 Background of the Invention

Several recent studies have demonstrated that nitric oxide has direct effects on myocardial contractile function. Exogenous nitric oxide donors characteristically enhance relaxation without major effects on peak systolic function. This selective relaxant action has been observed in papillary muscle preparations, intact hearts, and cardiac myocytes.

It has recently been determined that cyclic nucleotide-dependent relaxation of vascular smooth muscle is associated with an increase in the phosphorylation of the small heat shock related protein 20 ("HSP20"), and peptides derived from HSP20. HSP20 is highly and constitutively expressed in muscle tissues and can be phosphorylated in vitro by cGMP-dependent protein kinase.

However, the role of HSP20 and peptides derived therefrom in modulation of the contractile response in cardiac muscles is not known. Increasing the contractile rate in heart muscle would be of value in the prevention and treatment of various cardiac disorders.

25 Summary of the Invention

The present invention provides methods for increasing the contractile rate in heart muscle comprising administering to an individual in need thereof an amount effective to increase the contractile rate in heart muscle of one or more polypeptides comprising a sequence disclosed herein. In a preferred embodiment, increasing the contractile rate in heart muscle comprises increasing the rate of heart muscle

relaxation. In a further preferred embodiment, the individual to be treated suffers from one or more of bradyarrythmia, bradycardia, congestive heart failure, stunned myocardium, pulmonary hypertension, and diastolic dysfunction.

In another aspect, the present invention comprises methods for treating or preventing a cardiac disorder comprising administering to an individual suffering from one or more of bradyarrythmia, bradycardia, congestive heart failure, stunned myocardium, pulmonary hypertension, and diastolic dysfunction an amount effective to increase heart muscle contraction rate of one or more polypeptides comprising a sequence disclosed herein.

10 Detailed Description of the Invention

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In one aspect, the present invention provides methods for increasing the contractile rate in heart muscle comprising administering to an individual in need thereof an amount effective to increase the contractile rate in heart muscle of one or more polypeptides comprising a sequence according one or more of:

(a) general formula I:

 $X1-X2-[X3-A(X4)APLP-X5-]_u-X6$

wherein X1 is absent or is one or more molecules comprising one or more aromatic ring;

X2 is absent or comprises a transduction domain;

X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);

X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;

X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,

wherein Z1 is selected from the group consisting of G and D;

Z2 is selected from the group consisting of L and K; and

Z3 is selected from the group consisting of S, T, and K;

X6 is absent or comprises a transduction domain; and

wherein u is 1-5; and

(b) general formula II-:

30 $J1-J2-[J3-A(J4)APLP-J5]_{u}-J6$

wherein J1 is absent or is one or more molecules comprising one or more aromatic ring;

J2 is absent or comprises a cell transduction domain;

J3 is 0-14 amino acids of the sequence of heat shock protein 20 between residues 1 and 14 of SEQ ID NO: 298;

J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;

J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160 of SEQ ID NO:298; and

J6 is absent or comprises a cell transduction domain.

As used herein, an "individual in need thereof" means an individual that can benefit from an increased heart muscle contractile rate. Such individuals are those who exhibit a reduced heart rate relative to either a normal heart rate for the individual, or relative to a "normal" heart rate for a similarly situated individual.

As used herein, the phrase "increasing the contractile rate in heart muscle" means any increase in contractile rate that provides a therapeutic benefit to the patient. Such a therapeutic benefit can be achieved, for example, by increasing the contractile rate to make it closer to a normal contractile rate for the individual, a normal contractile rate for a similarly situated individual, or some other desired target contractile rate.

In a preferred embodiment, the methods of the invention result in an increase of at least 5% in the contractile rate of the patient in need of such treatment. In further preferred embodiments, the methods of the invention result in an increase of at least 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, and/or 50% in the contractile rate of the patient in need of such treatment.

In a preferred embodiment, increasing the contractile rate in heart muscle is accomplished by increasing the heart muscle relaxation rate (ie: if the muscles relax faster they beat faster). In a more preferred embodiment, the methods of the invention result in an increase of at least 5% in the heart muscle relaxation rate of the patient in need of such treatment. In further preferred embodiments, the methods of the invention result in an increase of at least 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, and/or 50% in the heart muscle relaxation rate of the patient in need of such treatment.

In a preferred embodiment of this aspect of the invention, the methods are performed to treat one or more heart cardiac disorders that can benefit from increasing the contractile rate in heart muscle. Such cardiac disorders include bradyarrhythmias,

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bradycardias congestive heart failure, pulmonary hypertension, stunned myocardium, and diastolic dysfunction.

As used herein, "bradyarrythmia" means an abnormal decrease of the rate of the heartbeat to less than 60 beats per minute, generally cased by a disturbance in the electrical impulses to the heart. A common cause of bradyarrythmias is coronary heart disease, which leads to the formation of atheromas that limit the flow of blood to the cardiac tissue, and thus the cardiac tissue becomes damaged. Bradyarrythmias due to coronary artery disease occur more frequently after myocardial infarction. Symptoms include, but are not limited to, loss of energy, weakness, syncope, and hypotension.

As used herein, "Congestive heart failure" means an inability of the heart to pump adequate supplies of blood throughout the body. Such heart failure can be due to a variety of conditions or disorders, including but not limited to hypertension, anemia, hyperthyroidism, heart valve defects including but not limited to aortic stenosis, aortic insufficiency, and tricuspid insufficiency; congenital heart defects including but not limited to coarctation of the aorta, septal defects, pulmonary stenosis, and tetralogy of Fallot; arrythmias, myocardial infarction, cardiomyopathy, pulmonary hypertension, and lung disease including but not limited to chronic bronchitis and emphysema. Symptoms of congestive heart failure include, but are not limited to, fatigue, breathing difficulty, pulmonary edema, and swelling of the ankles and legs.

As used herein, "Stunned myocardium" means heart muscle that is not functioning (pumping/beating) due to cardiac ischemia (lack of blood flow/oxygen to the vessels supplying the heat muscle).

As used herein, "Diastolic dysfunction" means an inability of the heart to fill with blood during diastole (the resting phase of heart contraction). This condition usually occurs in the setting of left ventricular hypertrophy. The heart muscle becomes enlarged and stiff such that it cannot fill adequately. Diastolic dysfunction can result in heart failure and inadequate heart function.

As used herein, "Pulmonary hypertension" means a disorder in which the blood pressure in the arteries supplying the lungs is abnormally high. Causes include, but are not limited to, inadequate supply of oxygen to the lungs, such as in chronic bronchitis and emphysema; pulmonary embolism, and intestinal pulmonary fibrosis. Symptoms and signs of pulmonary hypertension are often subtle and nonspecific. In

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the later stages, pulmonary hypertension leads to right heart failure that is associated with liver enlargement, enlargement of veins in the neck and generalized edema.

In a further aspect, the present invention provides methods for treating a heart muscle disorder comprising administering to an individual suffering from one or more of bradyarrythmia, bradycardia, congestive heart failure, stunned myocardium, pulmonary hypertension, and diastolic dysfunction an amount effective to increase heart muscle contractile rate of one or more polypeptides comprising or consisting of a sequence according to one or more of general formulas I and II.

As used herein, "treat" or "treating" means accomplishing one or more of the following: (a) reducing the severity of the disorder; (b) limiting or preventing development of symptoms characteristic of the disorder(s) being treated; (c) inhibiting worsening of symptoms characteristic of the disorder(s) being treated; (d) limiting or preventing recurrence of the disorder(s) in patients that have previously had the disorder(s); and (e) limiting or preventing recurrence of symptoms in patients that were previously symptomatic for the disorder(s).

For example, treating bradyarrythmia includes one or more of the following (a) improving the rate of the heartbeat to closer to normal levels for the individual, closer to a desired rate, or increasing to at least above 60 beats per minute; (b) preventing the occurrence of one or more of loss of energy, weakness, syncope, and hypotension in patients suffering from bradyarrythmia; (c) inhibiting worsening of one or more of loss of energy, weakness, syncope, and hypotension in patients suffering from bradyarrythmia and its symptoms; (d) limiting or preventing recurrence of bradyarrythmia in patients that previously suffered from bradyarrythmia; and (e) limiting or preventing recurrence of one or more of loss of energy, weakness, syncope, and hypotension in patients that previously suffered from bradyarrythmia.

Similarly, treating congestive heart failure includes one or more of the following (a) improving the heart's ability to pump adequate supplies of blood throughout the body to closer to normal levels for the individual, or closer to a desired pumping capacity; (b) limiting or preventing development of one or more of fatigue, breathing difficulty, pulmonary edema, and swelling of the ankles and legs in patients suffering from congestive heart failure; (c) inhibiting worsening of one or more of fatigue, breathing difficulty, pulmonary edema, and swelling of the ankles and legs in patients suffering from congestive heart failure and its symptoms; (d) limiting or

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preventing recurrence of congestive heart failure in patients that previously suffered from congestive heart failure; and (e) limiting or preventing recurrence of one or more of fatigue, breathing difficulty, pulmonary edema, and swelling of the ankles and legs in patients that previously suffered from congestive heart failure.

Treating stunned myocardium means one or more of (a) improving the ability of the heart muscle to pump by improving the oxygenation of the ischemic muscle, or by decreasing the need of the myocardial cells for oxygen and (b) limiting or preventing recurrence of stunned myocardium in patients that previously suffered from stunned myocardium.

Similarly, treating diastolic dysfunction includes one or more of (a) limiting or preventing heart failure and/or inadequate heart function by allowing the heart to relax and fill more completely; (b) limiting or preventing recurrence of diastolic dysfunction in patients that previously suffered from diastolic dysfunction; and (c) limiting or preventing recurrence of heart failure and/or inadequate heart function in patients that previously suffered from diastolic dysfunction.

Treating pulmonary hypertension includes one or more of the following (a) decreasing blood pressure in the arteries supplying the lungs to closer to normal levels for the individual, or closer to a desired pressure; (b) limiting or preventing the occurrence of one or more of enlargement of veins in the neck, enlargement of the liver, and generalized edema in patients suffering from pulmonary hypertension; (c) inhibiting worsening of one or more of enlargement of veins in the neck, enlargement of the liver, and generalized edema in patients suffering from pulmonary hypertension and its symptoms; (d) limiting or preventing recurrence of pulmonary hypertension in patients that previously suffered from pulmonary hypertension; and (e) limiting or preventing recurrence of one or more of enlargement of veins in the neck, enlargement of the liver, and generalized edema in patients that previously suffered from pulmonary hypertension.

In a further aspect, the present invention provides methods for preventing a heart muscle disorder comprising administering to an individual at risk of developing bradyarrythmia, bradycardia, congestive heart failure, stunned myocardium, pulmonary hypertension, and diastolic dysfunction an amount effective to increase heart muscle contractile rate of one or more polypeptides comprising or consisting of a sequence according to one or more of general formulas I and II.

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As used herein, the term "prevent" or "preventing" means to limit the disorder in individuals at risk of developing the disorder.

For example, methods to prevent congestive heart failure involve administration of the polypeptides as described above to a subject that suffers from one or more of hypertension, anemia, hyperthyroidism, heart valve defects including but not limited to aortic stenosis, aortic insufficiency, and tricuspid insufficiency; congenital heart defects including but not limited to coarctation of the aorta, septal defects, pulmonary stenosis, and tetralogy of Fallot; arrythmias, myocardial infarction, cardiomyopathy, pulmonary hypertension, and lung disease including but not limited to chronic bronchitis and emphysema.

Similarly, methods to prevent bradyarrythmia involve administration of the polypeptides as described above to a subject that suffer from one or more of coronary heart disease and atheroma formation, or that previously had a myocardial infarction or conduction disorder.

Similarly, methods to prevent pulmonary hypertension involve administration of the polypeptides as described above to a subject that suffers from one or more of chronic bronchitis, emphysema, pulmonary embolism, and intestinal pulmonary fibrosis.

Preventing stunned myocardium involves administration of the polypeptides as described above to a subject that suffers from cardiac ischemia.

Preventing, treating diastolic dysfunction involves administration of the polypeptides as described above to a subject that suffers from left ventricular hypertrophy

As used herein, an "amount effective" of the one or more polypeptides is an amount that is sufficient to provide the intended benefit of treatment. An effective amount of the polypeptides that can be employed ranges generally between about 0.01 μ g/kg body weight and about 10 mg/kg body weight, preferably ranging between about 0.05 μ g/kg and about 5 mg/kg body weight.

The term "polypeptide" is used in its broadest sense to refer to a sequence of subunit amino acids, amino acid analogs, or peptidomimetics. The subunits are linked by peptide bonds, except where noted (including when the X2 position is a non-amino acid molecule that contains an aromatic ring). The polypeptides described herein may be chemically synthesized or recombinantly expressed. Recombinant expression can be accomplished using standard methods in the art, generally involving the cloning of

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nucleic acid sequences capable of directing the expression of the polypeptides into an expression vector, which can be used to transfect or transduce a host cell in order to provide the cellular machinery to carry out expression of the polypeptides. Such expression vectors can comprise bacterial or viral expression vectors, and such host cells can be prokaryotic or eukaryotic.

Preferably, the polypeptides for use in the methods of the present invention are chemically synthesized. Synthetic polypeptides, prepared using the well-known techniques of solid phase, liquid phase, or peptide condensation techniques, or any combination thereof, can include natural and unnatural amino acids. Amino acids used for peptide synthesis may be standard Boc (Nα-amino protected Nα-t-butyloxycarbonyl) amino acid resin with the standard deprotecting, neutralization, coupling and wash protocols of the original solid phase procedure of Merrifield (13), or the base-labile Nα-amino protected 9-fluorenylmethoxycarbonyl (Fmoc) amino acids first described by Carpino and Han (5). Both Fmoc and Boc Nα-amino protected amino acids can be obtained from Sigma, Cambridge Research Biochemical, or other chemical companies familiar to those skilled in the art. In addition, the polypeptides can be synthesized with other Nα-protecting groups that are familiar to those skilled in this art.

Solid phase peptide synthesis may be accomplished by techniques familiar to those in the art and provided, for example, in (17, 7) or using automated synthesizers. The polypeptides of the invention may comprise D-amino acids (which are resistant to L-amino acid-specific proteases in vivo), a combination of D- and L-amino acids, and various "designer" amino acids (e.g., β -methyl amino acids, C α -methyl amino acids, and N α -methyl amino acids, etc.) to convey special properties. Synthetic amino acids include ornithine for lysine, and norleucine for leucine or isoleucine.

In addition, the polypeptides can have peptidomimetic bonds, such as ester bonds, to prepare polypeptides with novel properties. For example, a peptide may be generated that incorporates a reduced peptide bond, i.e., R_1 -CH₂-NH- R_2 , where R_1 and R_2 are amino acid residues or sequences. A reduced peptide bond may be introduced as a dipeptide subunit. Such a polypeptide would be resistant to protease activity, and would possess an extended half-live in vivo.

In a preferred embodiment of the polypeptides according to general formula I for use in the methods, X4 is phosphorylated. In a further preferred embodiment of

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the polypeptides according to general formula I, at least one of X2 and X6 comprises a transduction domain.

According to various embodiments of the polypeptides of general formula I, the region [X3-A(X4)APLP-X5-]_u may be present in 1, 2, 3, 4, or 5 copies. In a preferred embodiment, it is present in 1 copy. In other embodiments, it is present in multiple copies to provide increased efficacy for use in the methods of the invention.

According to various embodiments of the polypeptides of general formula I, X4 is S, T, Y, D E, a phosphoserine mimic, or a phosphotyrosine mimic. It is more preferred that X4 is S, T, or Y; more preferred that X4 is S or T, and most preferred that X4 is S. In these embodiments where X4 is S, T, or Y, it is most preferred that X4 is phosphorylated. When X4 is D or E, these residues have a negative charge that The polypeptides of general formula I are mimics the phosphorylated state. optimally effective in the methods of the invention when X4 is phosphorylated, is a phosphoserine or phosphotyrosine mimic, or is another mimic of a phosphorylated amino acid residue, such as a D or E residue. Examples of phosphoserine mimics include, but are not limited to, sulfoserine, amino acid mimics containing a methylene substitution for the phosphate oxygen, 4-phosphono(difluoromethyl)phenylanaline, and L-2-amino-4-(phosphono)-4,4-difuorobutanoic acid. Other phosphoserine mimics can be made by those of skill in the art; for example, see (15). Examples of limited but are not to, phosphotyrosine mimics include, phosphonomethylphenylalanine, difluorophosphonomethylphenylalanine, fluoro-Omalonyltyrosine and O-malonyltyrosine. (See, for example, (1)).

In another embodiment of the polypeptides of general formula I, X1 is one or more molecules comprising an aromatic ring. In one preferred embodiment, the one or molecules comprising an aromatic ring are amino acids, and X1 is $(F/Y/W)_z$, wherein "z" is 1-5 amino acids. Thus, for example, X1 can be 1 or 2 amino acid residues of any combination of F, Y, and W, such as F, FF, Y, YY, W, WW, FY, FW, YF, YW, WY, and WF. Alternatively, X1 can be a 3, 4, or 5 amino acid combination of F, Y, and W. In another preferred embodiment, the molecule comprising an aromatic ring is selected from the group of molecules comprising one or more aromatic rings which can optionally be substituted with halogen, lower alkyl, lower alkylthio, trifluoromethyl, lower acyloxy, aryl, and heteroaryl. In a most preferred embodiment, the one or more molecule comprising one or more aromatic ring comprise 9-fluorenylmethyl (Fm). Examples of such molecules include, but are not

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limited to 9-fluorenylmethylcarbonyl, 9-fluorenylmethylcarbamates, 9-fluorenylmethylcarbonates, 9-fluorenylmethyl esters, 9-fluorenylmethylphosphates, and S-9-fluorenylmethyl thioethers. In embodiments wherein the molecule comprising an aromatic ring is not an amino acid, it can be attached to the polypeptide by methods known in the art, including but not limited to, standard Fmoc protection chemistry employed in peptide synthesis.

According to other embodiments of the polypeptides of general formula I, X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1). If X3 consists of only one amino acid of the sequence, an "R" is present, since it is the carboxy-terminal amino acid of the sequence and it would be present at the amino terminus of the rest of the A(X4)APLP (SEQ ID NO: 2) sequence. If X3 consists of two amino acids of WLRR (SEQ ID NO:1), then the two amino acids added will be "RR". Other variations will be apparent to one of skill in the art based on the teachings herein. Similarly, variations in the residues that can make up X5 will be apparent to one of skill in the art based on the teachings herein.

Thus, according to these various embodiments, a representative sample of polypeptides according to general formula I for use in the methods of the invention include, but are not limited to, polypeptides comprising or consisting of the following ID NO:4); sequences: (ASAPLP)_u (SEQ ID NO:3); (ATAPLP)_u (SEQ (RASAPLP)_u (SEQ ID NO:5); (RATAPLP)_u (SEQ ID NO:6); (AYAPLP)_u (SEQ 20 ID NO:7); (RAYAPLP)_u (SEQ ID NO:8);(RRASAPLP)_u (SEQ ID NO:9); (WLRRASAPLP)_u; (SEQ ID NO:11) (LRRASAPLP), (SEQ ID NO:10); (RRATAPLP)_u (SEQ ID NO:12); (LRRATAPLP)_u (SEQ ID NO:13); (WLRRATAPLP)_u (SEQ ID NO:14); (RRAYAPLP)_u (SEQ ID NO:15); (LRRAYAPLP)_u (SEQ ID NO:16); (WLRRAYAPLP)_u (SEQ ID NO:17); 25 (RRASAPLPG)_u (SEQ ID NO:18); (RRASAPLPD)_u (SEQ ID NO:19); (RRASAPLPGL)_u (SEQ ID NO:20); (RRASAPLPGK)_u (SEQ ID NO:21); (RRASAPLPDL)_u (SEQ ID NO:22); (RRASAPLPDK)_u (SEQ ID NO:23); (RRASAPLPGLS)_u (SEQ ID NO:24); (RRASAPLPGLT)_u (SEQ ID NO:25); (RRASAPLPGKS), (SEQ ID NO:26); (RRASAPLPGKT), (SEQ ID NO:27); 30 (RRASAPLPDLS)_u (SEQ ID NO:28); RRASAPLPDLT)_u (SEQ ID NO:29); (RRASAPLPDKS)_u (SEQ ID NO:30); (RRASAPLPDKT)_u (SEQ ID NO:31); (LRRASAPLPG)_u (SEQ ID NO:32); (LRRASAPLPD)_u (SEQ ID NO:33); (LRRASAPLPGL)_u (SEQ ID NO:34); (LRRASAPLPGK)_u (SEQ ID NO:35);

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(LRRASAPLPDL)_u (SEQ ID NO:36); (LRRASAPLPDK)_u (SEQ ID NO:37); (LRRASAPLPGLS)_u (SEQ ID NO:38); (LRRASAPLPGLT)_u (SEQ ID NO:39); (LRRASAPLPGKS)_u (SEQ ID NO:40); (LRRASAPLPGKT)_u (SEQ ID NO:41); (LRRASAPLPDLS)₁₁ (SEQ ID NO:42); (LRRASAPLPDLT)₁₂ (SEQ ID NO:43); (LRRASAPLPDKS)_u (SEQ ID NO:44); (LRRASAPLPDKT)_u (SEQ ID NO:45); 5 (WLRRASAPLPG)_u (SEQ ID NO:46); (WLRRASAPLPD)_u (SEQ ID NO:47); (WLRRASAPLPGL)_u (SEQ ID NO:48); (WLRRASAPLPGK)_u (SEQ ID NO:49); (WLRRASAPLPDL)_u (SEQ ID NO:50); (WLRRASAPLPDK)_u (SEQ ID NO:51); (WLRRASAPLPGLS)_u (SEQ ID NO:52); (WLRRASAPLPGLT)_u (SEQ ID NO:53); (WLRRASAPLPGKS), (SEQ ID NO:54); (WLRRASAPLPGKT), (SEQ ID 10 NO:55); (WLRRASAPLPDLS)u (SEQ ID NO:56); (WLRRASAPLPDLT)u (SEQ ID NO:57); (WLRRASAPLPDKS)_u (SEQ ID NO:58); (WLRRASAPLPDKT)_u (SEQ ID NO:59); (RRATAPLPG)_u (SEQ ID NO:60); (RRATAPLPD)_u (SEQ ID NO:61); (RRATAPLPGL)_u (SEQ ID NO:62); (RRATAPLPGK)_u (SEQ ID NO:63); 15 (RRATAPLPDL)_u (SEQ ID NO:64); (RRATAPLPDK)_u (SEQ ID NO:65); (RRATAPLPGLS)_u (SEQ ID NO:66); (RRATAPLPGLT)_u (SEQ ID NO:67); (RRATAPLPGKS)_u (SEQ ID NO:68); (RRATAPLPGKT)_u (SEQ ID NO:69); (RRATAPLPDLS), (SEQ ID NO:70); (RRATAPLPDLT), (SEQ ID NO:71); (RRATAPLPDKS)_u (SEQ ID NO:72); (RRATAPLPDKT)_u (SEQ ID NO:73); $(LRRATAPLPG)_u$ (SEQ ID NO:74); $(LRRATAPLPD)_u$ (SEQ ID NO:75); 20 (LRRATAPLPGL)_u (SEQ ID NO:76); (LRRATAPLPGK)_u (SEQ ID NO:77); (LRRATAPLPDL)_u (SEQ ID NO:78); (LRRATAPLPDK)_u (SEQ ID NO:79); (LRRATAPLPGLS)_u (SEQ ID NO:80); (LRRATAPLPGLT)_u (SEQ ID NO:81); (LRRATAPLPGKS), (SEQ ID NO:82); (LRRATAPLPGKT), (SEQ ID NO:83); (LRRATAPLPDLS)_u (SEQ ID NO:84); (LRRATAPLPDLT)_u (SEQ ID NO:85); 25 (LRRATAPLPDKS)_u (SEQ ID NO:86); (LRRATAPLPDKT)_u (SEQ ID NO:87); (WLRRATAPLPG)_u (SEQ ID NO:88); (WLRRATAPLPD)_u (SEQ ID NO:89); (WLRRATAPLPGL)_u (SEQ ID NO:90); (WLRRATAPLPGK)_u (SEQ ID NO:91); (WLRRATAPLPDL)_u (SEQ ID NO:92); (WLRRATAPLPDK)_u (SEQ ID NO:93); (WLRRATAPLPGLS)_u (SEQ ID NO:94); (WLRRATAPLPGLT)_u (SEQ ID 30 NO:95); (WLRRATAPLPGKS)_u (SEQ ID NO:96); (WLRRATAPLPGKT)_u (SEQ ID NO:97); (WLRRATAPLPDLS)_u (SEQ ID NO:98); (WLRRATAPLPDLT)_u (SEQ ID NO:100); (WLRRATAPLPDKS)_u (SEO ID NO:99); (WLRRATAPLPDKT)_u (SEQ ID NO:101); (RRAYAPLPG)_u (SEQ ID NO:102);

(RRAYAPLPD), (SEQ ID NO:103); (RRAYAPLPGL), (SEQ ID NO:104); (RRAYAPLPGK)_u (SEQ ID NO:105); (RRAYAPLPDL)_u (SEQ ID NO:106); (RRAYAPLPDK)_u (SEQ ID NO:107); (RRAYAPLPGLS)_u (SEQ ID NO:108); (RRAYAPLPGLT)_u (SEQ ID NO:109); (RRAYAPLPGKS)_u (SEQ ID NO:110; (RRAYAPLPGKT)_u (SEQ ID NO:111); (RRAYAPLPDLS)_u (SEQ ID NO:112); 5 (RRAYAPLPDLT)_u (SEQ ID NO:113); (RRAYAPLPDKS)_u (SEQ ID NO:114); (RRAYAPLPDKT)_u (SEQ ID NO:115); (LRRAYAPLPG)_u (SEQ ID NO:116); (LRRAYAPLPD)_u (SEQ ID NO:117); (LRRAYAPLPGL)_u (SEQ ID NO:118); (LRRAYAPLPGK)_u (SEQ ID NO:119); (LRRAYAPLPDL)_u (SEQ ID NO:120); (LRRAYAPLPDK)_u (SEQ ID NO:121); (LRRAYAPLPGLS)_u (SEQ ID NO:122); 10 (LRRAYAPLPGLT)_u (SEQ ID NO:123); (LRRAYAPLPGKS)_u (SEQ ID NO:124); (LRRAYAPLPGKT)_u (SEQ ID NO:125); (LRRAYAPLPDLS)_u (SEQ ID NO:126); (LRRAYAPLPDLT), (SEQ ID NO:127); (LRRAYAPLPDKS), (SEQ ID NO:128); (LRRAYAPLPDKT)_u (SEQ ID NO:129); (WLRRAYAPLPG)_u (SEQ ID NO:130); (WLRRAYAPLPD)_u (SEQ ID NO:131); (WLRRAYAPLPGL)_u (SEQ ID NO:132); 15 (WLRRAYAPLPGK)_u (SEQ ID NO:133); (WLRRAYAPLPDL)_u (SEQ ID NO:134); (WLRRAYAPLPDK)_u (SEQ ID NO:135); (WLRRAYAPLPGLS)_u (SEQ ID NO:136); (WLRRAYAPLPGLT)_u (SEQ ID NO:137); (WLRRAYAPLPGKS)_u (WLRRAYAPLPGKT)_u \mathbf{m} NO:139); ID NO:138); (SEQ (SEQ (WLRRAYAPLPDLS)_u (SEQ ID NO:140); (WLRRAYAPLPDLT)_u (SEQ ID 20 NO:141); (WLRRAYAPLPDKS)_u (SEQ ID NO:142); and (WLRRAYAPLPDKT)_u ID NO:144); ((F/Y/W)RRASAPLP)_u (SEQ NO:143); (SEQ \mathbf{ID} ((F/Y/W)LRRASAPLP)_u (SEQ ID NO:145); ((F/Y/W)WLRRASAPLP)_u; (SEQ ID NO:146) ((F/Y/W)RRATAPLP)_u (SEQ ID NO:147); ((F/Y/W)LRRATAPLP)_u (SEQ ((F/Y/W)WLRRATAPLP)_u (SEQ \mathbf{m} NO:149); 25 \mathbf{ID} NO:148); ((F/Y/W)RRAYAPLP)_u (SEQ ID NO:150); ((F/Y/W)LRRAYAPLP)_u (SEQ ID NO:151); ((F/Y/W)WLRRAYAPLP)_u (SEQ ID NO:152); ((F/Y/W)RRASAPLPG)_u NO:154); (SEQ \mathbf{ID} NO:153); ((F/Y/W)RRASAPLPD)_u (SEQ ID ((F/Y/W)RRASAPLPGL)_u (SEQ ID NO:155); ((F/Y/W)RRASAPLPGK)_u (SEQ ID NO:156); ((F/Y/W)RRASAPLPDL)_u (SEQ ID NO:157); ((F/Y/W)RRASAPLPDK)_u 30 ((F/Y/W)RRASAPLPGLS)_u (SEQ \mathbf{m} NO:159); NO:158); (SEO ID ((F/Y/W)RRASAPLPGLT)_u (SEQ ID NO:160); ((F/Y/W)RRASAPLPGKS)_u; (SEQ \mathbf{D} NO:162); (SEQ ((F/Y/W)RRASAPLPGKT)_u ID NO:161); ((F/Y/W)RRASAPLPDLS)_u (SEQ ID NO:163); ((F/Y/W)RRASAPLPDLT)_u (SEQ

 \mathbf{m} (SEQ NO:165); ((F/Y/W)RRASAPLPDKS)_u ID NO:164); ((F/Y/W)RRASAPLPDKT)_u (SEQ ID NO:166); ((F/Y/W)LRRASAPLPG)_u (SEQ ((F/Y/W)LRRASAPLPD)_u (SEQ \mathbf{m} NO:168); ID NO:167); ((F/Y/W))LRRASAPLPGL)₁₁ (SEQ ID NO:169); ((F/Y/W)LRRASAPLPGK)₁₁ (SEQ ((F/Y/W)LRRASAPLPDL)_u (SEQ ID NO:171); 5 \mathbf{m} NO:170); ((F/Y/W)LRRASAPLPDK)_u (SEQ ID NO:172); ((F/Y/W)LRRASAPLPGLS)_u (SEQ \mathbf{D} NO:173); ((F/Y/W)LRRASAPLPGLT)_u (SEQ ID NO:174); ((F/Y/W)LRRASAPLPGKS), (SEQ ID NO:175); ((F/Y/W)LRRASAPLPGKT), ((F/Y/W)LRRASAPLPDLS)_u (SEQ ID (SEQ ID NO:176); ((F/Y/W)LRRASAPLPDLT)_u (SEQ ID NO:178); ((F/Y/W)LRRASAPLPDKS)_u 10 (SEQ ID NO:179); ((F/Y/W)LRRASAPLPDKT)_u (SEQ ID ((F/Y/W)WLRRASAPLPG)_u (SEQ ID NO:181); ((F/Y/W)WLRRASAPLPD)_u (SEQ ((F/Y/W)WLRRASAPLPGL)_u (SEQ \mathbf{m} NO:183); NO:182); ((F/Y/W)WLRRASAPLPGK)_u (SEQ ID NO:184); ((F/Y/W)WLRRASAPLPDL)_u (SEO ID NO:185); ((F/Y/W)WLRRASAPLPDK)_u (SEQ ID NO:186); 15 ((F/Y/W)WLRRASAPLPGLS)₁₁ (SEQ ID NO:187); ((F/Y/W)WLRRASAPLPGLT)₁₁ (SEQ ID NO:188); ((F/Y/W)WLRRASAPLPGKS)_u (SEQ ID NO:189); ((F/Y/W)WLRRASAPLPGKT), (SEQ ID NO:190); ((F/Y/W)WLRRASAPLPDLS), (SEQ ID NO:191); ((F/Y/W)WLRRASAPLPDLT)_u (SEQ ID NO:192); ((F/Y/W)WLRRASAPLPDKS)_u (SEQ ID NO:193); ((F/Y/W)WLRRASAPLPDKT)_u 20 ((F/Y/W)RRATAPLPG)_u (SEQ \mathbf{ID} NO:195); NO:194); (SEQ ID ((F/Y/W)RRATAPLPD)_u (SEQ ID NO:196); ((F/Y/W)RRATAPLPGL)_u (SEQ ID NO:197); ((F/Y/W)RRATAPLPGK), (SEQ ID NO:198); ((F/Y/W)RRATAPLPDL), ((F/Y/W)RRATAPLPDK)_u (SEQ ID NO:200); NO:199); (SEQ ((F/Y/W)RRATAPLPGLS)_u (SEQ ID NO:201); ((F/Y/W)RRATAPLPGLT)_u (SEQ 25 ((F/Y/W)RRATAPLPGKS)_u (SEQ ID NO:203); NO:202); ((F/Y/W)RRATAPLPGKT)_u (SEQ ID NO:204); ((F/Y/W)RRATAPLPDLS)_u (SEQ (SEQ ID NO:206); \mathbf{ID} NO:205): ((F/Y/W)RRATAPLPDLT)_u ((F/Y/W)RRATAPLPDKS)_u (SEQ ID NO:207); ((F/Y/W)RRATAPLPDKT)_u (SEQ ((F/Y/W)LRRATAPLPG)_u (SEQ \mathbf{ID} NO:209); 30 \mathbf{ID} NO:208); ((F/Y/W)LRRATAPLPD)_u (SEQ ID NO:210); ((F/Y/W)LRRATAPLPGL)_u (SEQ \mathbf{m} NO:212); ((F/Y/W)LRRATAPLPGK)_u (SEQ \mathbf{m} NO:211); ((F/Y/W)LRRATAPLPDL)_u (SEQ ID NO:213); ((F/Y/W)LRRATAPLPDK)_u (SEQ \mathbf{m} NO:215); \mathbf{m} NO:214); ((F/Y/W)LRRATAPLPGLS), (SEQ

((F/Y/W)LRRATAPLPGLT)_u (SEQ ID NO:216); ((F/Y/W)LRRATAPLPGKS)_u (SEO ID NO:217); ((F/Y/W)LRRATAPLPGKT), (SEQ ID NO:218); ((F/Y/W)LRRATAPLPDLS)_u (SEQ ID NO:219); ((F/Y/W)LRRATAPLPDLT)_u ID NO:220); $((F/Y/W)LRRATAPLPDKS)_u$ (SEQ ID NO:221); (SEO ((F/Y/W)LRRATAPLPDKT)_u (SEQ ID NO:222); ((F/Y/W)WLRRATAPLPG)_u (SEQ \mathbf{m} NO:223); $((F/Y/W)WLRRATAPLPD)_u$ (SEQ ${f D}$ ((F/Y/W)WLRRATAPLPGL)_u (SEQ ID NO:225); ((F/Y/W)WLRRATAPLPGK)_u (SEQ ID NO:226); ((F/Y/W)WLRRATAPLPDL)_u (SEQ ID NO:227); ((F/Y/W)WLRRATAPLPDK)_u (SEQ ID NO:228); ((F/Y/W)WLRRATAPLPGLS)_u (SEQ ID NO:229); ((F/Y/W)WLRRATAPLPGLT)_u (SEQ ID NO:230); 10 ((F/Y/W)WLRRATAPLPGKS)_u (SEQ \mathbf{m} NO:231); ((F/Y/W)WLRRATAPLPGKT)_u (SEQ ID NO:232); ((F/Y/W)WLRRATAPLPDLS)_u (SEQ ID NO:233); ((F/Y/W)WLRRATAPLPDLT)_u (SEQ ID NO:234); \mathbf{ID} NO:235); ((F/Y/W)WLRRATAPLPDKS)_u (SEQ ((F/Y/W)WLRRATAPLPDKT)_u (SEQ ID NO:236); ((F/Y/W)RRAYAPLPG)_u (SEQ 15 ID NO:237); ((F/Y/W)RRAYAPLPD)_u (SEQ \mathbf{ID} NO:238); ((F/Y/W)RRAYAPLPGL)_u (SEQ ID NO:239); ((F/Y/W)RRAYAPLPGK)_u (SEQ ID \mathbf{m} NO:241); ((F/Y/W)RRAYAPLPDL)_u (SEQ NO:240); ((F/Y/W)RRAYAPLPDK)u (SEQ ID NO:242); ((F/Y/W)RRAYAPLPGLS)u. (SEQ \mathbf{D} 20 \mathbf{ID} NO:243); ((F/Y/W)RRAYAPLPGLT)_u (SEQ NO:244); ((F/Y/W)RRAYAPLPGKS)₁₁ (SEQ ID NO:245); ((F/Y/W)RRAYAPLPGKT)₁₂ (SEQ ID ((F/Y/W)RRAYAPLPDLS)_u (SEQ NO:247); ID NO:246); ((F/Y/W)RRAYAPLPDLT)_u (SEQ ID NO:248); ((F/Y/W)RRAYAPLPDKS)_u (SEQ ID ((F/Y/W)RRAYAPLPDKT)_u (SEQ ID NO:250); NO:249); ((F/Y/W)LRRAYAPLPG)_u (SEQ ID NO:251); ((F/Y/W)LRRAYAPLPD)_u (SEQ ID 25 ((F/Y/W)LRRAYAPLPGL)_u (SEQ \mathbf{ID} NO:253); NO:252); ((F/Y/W)LRRAYAPLPGK)_u (SEQ ID NO:254); ((F/Y/W)LRRAYAPLPDL)_u (SEQ ((F/Y/W)LRRAYAPLPDK)_u (SEQ ID NO:256); NO:255): ((F/Y/W)LRRAYAPLPGLS)_u (SEQ ID NO:257); ((F/Y/W)LRRAYAPLPGLT)_u (SEQ ID NO:258); ((F/Y/W)LRRAYAPLPGKS)_u (SEQ ID 30 ((F/Y/W)LRRAYAPLPGKT)_u (SEQ ID NO:260); ((F/Y/W)LRRAYAPLPDLS)_u (SEQ ID NO:261); ((F/Y/W)LRRAYAPLPDLT)_u (SEQ ID NO:262); ((F/Y/W)LRRAYAPLPDKS)_u (SEQ ID NO:263); ((F/Y/W)LRRAYAPLPDKT)_u (SEQ ID NO:264); ((F/Y/W)WLRRAYAPLPG)_u (SEQ ID

((F/Y/W)WLRRAYAPLPD)_u (SEQ ID NO:266); ((F/Y/W)WLRRAYAPLPGL)_u NO:268); (SEQ ID NO:267); ((F/Y/W)WLRRAYAPLPGK)_u (SEQ ((F/Y/W)WLRRAYAPLPDL)_u (SEQ ID NO:269); ((F/Y/W)WLRRAYAPLPDK)_u (SEQ ID NO:270); ((F/Y/W)WLRRAYAPLPGLS)_u (SEQ \mathbf{ID} NO:271); NO:272); ID ((F/Y/W)WLRRAYAPLPGLT)_u (SEQ NO:273); (SEQ \mathbf{m} ((F/Y/W)WLRRAYAPLPGKS)_u ((F/Y/W)WLRRAYAPLPGKT)u (SEQ \mathbf{m} NO:274); (SEQ \mathbf{m} NO:275); ((F/Y/W)WLRRAYAPLPDLS)_u ((F/Y/W)WLRRAYAPLPDLT),, (SEQ ID NO:276); \mathbf{ID} NO:277); and ((F/Y/W)WLRRAYAPLPDKS)_u (SEQ 10 ((F/Y/W)WLRRAYAPLPDKT)_u (SEQ ID NO:278) wherein "u" is as defined above, and (F/Y/W) means that the residue is selected from F, Y, and W. Other specific polypeptides falling within the scope of general formula I will be readily apparent to one of skill in the art based on the teachings herein.

In a further embodiment, the polypeptides according to general formula I comprise a combination of different sequences from the region [X3-A (X4)APLP-X5l₁₁. In this embodiment, for example, the polypeptide can consist of 1 copy of SEQ ID NO: 9 and 1 copy of SEO ID NO: 143. In a different example, the polypeptide could consist of 2 copies of SEQID NO: 200 and 3 copies of SEQ ID NO: 62. It will be apparent to one of skill in the art that many such combinations are possible based on the teachings of the present invention.

In a preferred embodiment of the polypeptides according to general formula I, at least one of X2 and X6 comprises or consists of a transduction domain. As used herein, the term "transduction domain" means one or more amino acid sequences that can carry the active domain across cell membranes. These domains can be linked to other polypeptides to direct movement of the linked polypeptide across cell membranes. In some cases the transducing molecules do not need to be covalently linked to the active polypeptide (for example, see sequence ID 291). In a preferred embodiment, the transduction domain is linked to the rest of the polypeptide via peptide bonding. (See, for example, (9, 8, 6, 16, 19, 10)). In this embodiment, any of the polypeptides as described above would include at least one transduction domain. In a further embodiment, both X2 and X6 comprise transduction domains. In a further preferred embodiment, the transduction domain(s) is/are selected from the group consisting of (R)4-9 (SEQ ID NO:279); GRKKRRQRRRPPQ (SEQ ID

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NO:280); YARAAARQARA (SEQ ID NO:281);
DAATATRGRSAASRPTERPRAPARSASRPRRPVE (SEQ ID NO:282);
GWTLNSAGYLLGLINLKALAALAKKIL (SEQ ID NO:283); PLSSIFSRIGDP
(SEQ ID NO:284); AAVALLPAVLLALLAP (SEQ ID NO:285);

(SEQ ID NO:284); AAVALLPAVLLALLAP (SEQ ID NO:285);

5 AAVLLPVLLAAP (SEQ ID NO:286); VTVLALGALAGVGVG (SEQ ID NO:287); GALFLGWLGAAGSTMGAWSQP (SEQ ID NO:288);

GWTLNSAGYLLGLINLKALAALAKKIL (SEQ ID NO:289);

KLALKLALKALKAALKLA (SEQ ID NO:290);

KETWWETWWTEWSQPKKKRKV (SEQ ID NO:291); KAFAKLAARLYRKAGC

10 (SEQ ID NO:292); KAFAKLAARLYRAAGC (SEQ ID NO:293);

AAFAKLAARLYRKAGC (SEQ ID NO:294); KAFAALAARLYRKAGC (SEQ ID NO:295); KAFAKLAAQLYRKAGC (SEQ ID NO:296), and

GGGGYGRKKRRQRRR (SEQ ID NO:297).

The polypeptides of the invention according to general formula II recite that J4 is S, T, Y, D E, a phosphoserine mimic, or a phosphotyrosine mimic. It is preferred 15 that J4 is S, T, or Y; more preferred that J4 is S or T, and most preferred that J4 is S. In these embodiments where J4 is S, T, or Y, it is most preferred that J4 is phosphorylated. When J4 is D or E, these residues have a negative charge that mimics the phosphorylated state. The polypeptides according to general formula II are optimally effective in the methods of the invention when J4 is phosphorylated, is a 20 phosphoserine or phosphotyrosine mimic, or is another mimic of a phosphorylated amino acid residue, such as a D or E residue. Examples of phosphoserine mimics include, but are not limited to, sulfoserine, amino acid mimics containing a methylene substitution for the phosphate oxygen, 4-phosphono(difluoromethyl)phenylanaline, 25 and L-2-amino-4-(phosphono)-4,4-difuorobutanoic acid. Other phosphoserine mimics can be made by those of skill in the art; for example, see (15). Examples of phosphotyrosine mimics include, but are not limited to, phosphonomethylphenylalanine, difluorophosphonomethylphenylalanine, fluoro-Omalonyltyrosine and O-malonyltyrosine. (See, for example, (1)).

In a further preferred embodiment of the polypeptides according to general formula II, at least one of J2 and J6 comprises or consists of a cell transduction domain. In a further preferred embodiment, the carboxy-terminal end of the polypeptides according to general formula II are unblocked. In a further preferred

embodiment, the amino terminal end of the polypeptides according to general formula II are unblocked.

The HSP20 polypeptide sequence is as follows (SEQ ID NO: 298):

Met Glu Ile Pro Val Pro Val Gln Pro Ser Trp Leu Arg Arg Ala Ser Ala Pro Leu Pro

Gly Leu Ser Ala Pro Gly Arg Leu Phe Asp Gln Arg Phe Gly Glu Gly Leu Leu Glu

Ala Glu Leu Ala Ala Leu Cys Pro Thr Thr Leu Ala Pro Tyr Tyr Leu Arg Ala Pro Ser

Val Ala Leu Pro Val Ala Gln Val Pro Thr Asp Pro Gly His Phe Ser Val Leu Leu Asp

Val Lys His Phe Ser Pro Glu Glu Ile Ala Val Lys Val Val Gly Glu His Val Glu Val

His Ala Arg His Glu Glu Arg Pro Asp Glu His Gly Phe Val Ala Arg Glu Phe His Arg

Arg Tyr Arg Leu Pro Pro Gly Val Asp Pro Ala Ala Val Thr Ser Ala Leu Ser Pro Glu

Gly Val Leu Ser Ile Gln Ala Ala Pro Ala Ser Ala Gln Ala Pro Pro Pro Pro Ala Ala Ala

Lys.

The polypeptides may be subjected to conventional pharmaceutical operations such as sterilization and/or may contain conventional adjuvants, such as preservatives, stabilizers, wetting agents, emulsifiers, buffers etc.

For administration, the polypeptides are ordinarily combined with one or more adjuvants appropriate for the indicated route of administration. The compounds may be admixed with lactose, sucrose, starch powder, cellulose esters of alkanoic acids, stearic acid, talc, magnesium stearate, magnesium oxide, sodium and calcium salts of phosphoric and sulphuric acids, acacia, gelatin, sodium alginate, polyvinylpyrrolidine, dextran sulfate, heparin-containing gels, and/or polyvinyl alcohol, and tableted or encapsulated for conventional administration. Alternatively, the compounds of this invention may be dissolved in saline, water, polyethylene glycol, propylene glycol, carboxymethyl cellulose colloidal solutions, ethanol, corn oil, peanut oil, cottonseed oil, sesame oil, tragacanth gum, and/or various buffers. Other adjuvants and modes of administration are well known in the pharmaceutical art. The carrier or diluent may include time delay material, such as glyceryl monostearate or glyceryl distearate alone or with a wax, or other materials well known in the art.

The polypeptides or pharmaceutical compositions thereof may be administered by any suitable route, including orally, parentally, by inhalation spray, rectally, or topically in dosage unit formulations containing conventional pharmaceutically acceptable carriers, adjuvants, and vehicles. The term parenteral as used herein

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includes, subcutaneous, intravenous, intra-arterial, intramuscular, intrasternal, intratendinous, intraspinal, intracranial, intrathoracic, infusion techniques or intraperitoneally. Preferred embodiments for administration vary with respect to the condition being treated, and are described in detail below.

The polypeptides may be made up in a solid form (including granules, powders or suppositories) or in a liquid form (e.g., solutions, suspensions, or emulsions). The polypeptides of the invention may be applied in a variety of solutions. Suitable solutions for use in accordance with the invention are sterile, dissolve sufficient amounts of the polypeptides, and are not harmful for the proposed application.

In Experiments described herein it is demonstrated that treatment of neonatal ventricular tissue with sodium nitroprusside leads to increases in the rate of contraction and increased phosphorylation of HSP20. The addition of phosphopeptide analogues of HSP20 into transiently permeabilized cardiac myocytes led to increases in the rate of contraction of the myocytes. These results are consistent with previously reported findings demonstrating that the NO-cGMP pathway increases beat rate by stimulating the hyperpolarization-activated pacemaker current If (14). While not being bound by any specific mechanism of action, the present results suggest that the mechanism of NO stimulation of heart rate via If involves phosphorylation of HSP20 and that phosphorylated HSP20 may have a direct effect on If channels. The increased rate of contraction was also related to increases in the rate of relaxation of the myocytes. Consistent with this increase, is an increase in the rate of decline of the Ca²⁺ transient. While not being bound by any specific mechanism of action, these data suggest that phosphorylated HSP20 facilitates increased beat rate by stimulating a more rapid uptake of Ca²⁺ by the SR, and further suggest that HSP20 may have a direct role in modulating the lusitropic actions of nitric oxide and nitric oxide donors.

HSP20 is biochemically associated with αB -crystallin and co-localizes with αB -crystallin and sarcomeric actin at the I-band. The association of HSP20 and αB -crystallin with the I-band suggests that these two small heat shock proteins may be involved in modulating cytoskeletal and/or contractile dynamics of cardiac myocytes.

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Examples

Example 1

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Sodium dodecylsulfate (SDS); glycine; and tris-(hydroxymethyl) Materials: aminomethane (Tris); dithiothreitol (DTT); were from Research Organics (Cleveland, OR). Coomassie brilliant blue was from ICN Biomedicals Inc (Aurora, OR). Recombinant HSP27 was from StressGen (Victoria, BC Canada). Recombinant HSP20 was produced as previously described14. Piperazine diacrylamide and other electrophoresis/ reagents were from Biorad (Hercules, CA). 3-((3-cholamidopropyl dimethyl ammonio)-1-propanesulfonate (CHAPS); ethylene glycol bis (B-aminoethyl ether) - N, N, N' - tetra acetic acid (EGTA); ethylene diaminetetraacetic acid (EDTA); polyoxyethylene-sorbitan monolaurate (Tween-20); purified bovine aB-crystallin; sodium nitroprusside and all other reagent grade chemicals were from Sigma (St. Louis, MO). Molecular weight standards were from Pharmacia (Upsalla, Sweden). Immobilon was from Millipore (Bedford Ma). Antibodies against sarcomeric actin were from Sigma (St. Louis, Mo), HSP20 and 3αB-crystallin from Dr. Kanefusa Kato (Aichi, Japan) (12), HSP25 from Dr Michael Welsh (University of Michigan, Ann Arbor Michigan) (20) Accurate (Westbury, NY.), Stressgen (Victoria, BC) and myotonic dystrophy kinase binding protein from Dr. Atshushi Suzuki (Yokohama, Japan) (18). Goat anti-mouse and anti-rabbit secondary antibodies were from Jackson Immunochemical (West Grove, PA). Protein concentrations were determined using the Coomassie Plus Protein Assay Reagent (Pierce Rockford IL).

Isolation of rat heart tissue: Adult rats (2-3 months of age) were sacrificed with CO₂ inhalation and the hearts were dissected free from the thoracic cavity and placed in phosphate buffered saline (PBS, 10 mM phosphate, pH 7.5, 0.15 M NaCl, 4°C). The atria were dissected free and the ventricle was, cut into small strips, and homogenized for biochemical analyses. For immunohistochemical analyses, the thoracic aorta was cannulated and perfused with PBS followed by 4% formalin in PBS at 100 mmHg. Ventricular myocytes were isolated from the hearts of 2 day old Sprague-Dawley rats by gentle trypsinization and placed in culture as previously described (11). Animal use was approved by the Institutional Animal Care and Use Committee.

Immunoblotting: Tissues were homogenized in 10 mM EGTA, 2 mM EDTA, 10 mM β-mercaptoethanol, 1 % Glycerol, and 4% SDS in 60 mM Tris, pH 7.0 followed

by centrifugation (10 000 x g) to remove insoluble material. Proteins were separated on 15% SDS-PAGE gels and transferred to Immobilon for 210volt hours. The blots were air dried and subsequently blocked with tris buffered saline (TBS: 10 mM Tris, 150 mM NaCl pH 7.4)/5% milk for 1 hour. The blots were then incubated with anti-HSP20 (1-/1000), anti-αB-crystallin (1/1000), anti-HSP25 (1/1000) or anti-MKBP (1/1000) antibodies in TBS/milk for one hour at room temperature. The blots were washed 3 times (5 minutes each) in TBS/Tween-20 (0. 5%). The blots were then placed in goat anti-rabbit secondary antibody diluted in TBS/milk (1/2000) for one hour at room temperature. The blots were then washed 6 times (5 minutes each) in TBS/Tween-20. Immunoreactive protein was determined using enhanced chemiluminescence (DuPont NEN, Boston, MA) exposed on X-ray film.

Tissues were homogenized in homogenization buffer Subcellular fractionation: ("HEPES buffer" 25 mM HEPES, 150 mM NaCl, 10 mM EDTA, 1 mM DTT, 2 mM benzamidine, pH 7.4 (0. 5 gm tissue/1 ml buffer)) in a polytron homogenizer at 4°C. The homogenate was centrifuged 3000 x g to remove debris and the nuclear pellet. The supernatants were diluted to 5 µg of protein/µl and 250 µl of supernatant was centrifuged 10,000 x g for 10 minutes. The pellet (P 1) was resuspended in 125 µl of homogenization buffer and 125 µl of 2X sample buffer (6.25 mM Tris pH 6.8, 2% SDS, 5% 2- β -mercaptoethanol, 10% glycerol, 0.025% bromophenol blue). To 125 μ l of the 10,000 x g supernatant (S1), 125 µl of 2X sample buffer was added. The remaining 125 µl of 10,000 x g supernatant was centrifuged 100,000 x g. The 100,000 x g pellet (P2) was resuspended in 125 μl of homogenization buffer and 125 μl of 2X sample buffer. 125 μ l of 2X sample buffer was added to the 100,000 x g supernatant (S2). The samples were boiled for 8 minutes after the addition of sample buffer, separated on 15% SDS-PAGE gels, transferred to Immobilon membranes and probed with anti-actin antibodies (1/2000 dilution), anti-HSP20 antibodies (1/2000 dilution), anti-αB-crystallin antibodies (1/2000 dilution) and anti-MKBP antibodies (1/2000 dilution).

Gel Filtration: Gel filtration was performed as previously described (3). In brief, the tissues were homogenized in HEPES buffer (0. 5 gm tissue/1 ml buffer) at 4°C using a polytron homogenizer (Brinkman Instruments, Westbury, NY). The homogenate was centrifuged 100,000 X g for 30 minutes at 4°C. 200 μl of

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supernatant (containing 200 µg total protein) was applied to a Superose- HR 10/30 fast protein liquid chromatography column (Pharmacia, Upsalla Sweden), eluted with column buffer and 0.5 ml fractions collected. For calibration, 10 µl of catalase (232 kDa, 5 mg/ml) and bovine serum albumin (67 kDa, 8 mg/ml) were applied to the column.

Dot blotting: 100 μl of each fraction from the column was dot blotted onto nitrocellulose. The blots were fixed with 20% methanol, dried, blocked with TBS, 5% milk, for 1 hour, washed 3 times with TBS, and then probed with anti-HSP20, anti-MKBP, and anti-αB-crystallin antibodies (1/2000 dilution in TBS, 5% milk) for 1 hour. Goat anti-rabbit secondary antibodies (1/2000 dilution) were added to the blots for one hour. The blots were then washed 6 times with TBS/0.5% Tween-20. Immunoreactive proteins were visualized as described above with enhanced chemiluminescence and densitometric analysis was performed using UN-SCAN-IT automated digitizing software (Silk Scientific Corporation, Orem, UT).

15 Immunohistochemistry: Perfusion fixed ventricular tissue was embedded in paraffin. Five micron cross-sections were mounted on polylysine slides. The slides were de-paraffinized with xylene and graded dilutions of ethanol. The sections were rinsed in PBS and blocked with donkey serum (Jackson Immunoresearch, West Grove, PA.) for 30 minutes. The slides were then incubated with anti-HSP20 (1/100), anti-αB-crystallin (1/100), anti- sarcomeric actin (1/100), or anti-MKBP (1/100) overnight at 4°C. The slides were then washed with PBS, 4 times (15 minutes each), at room temperature. Anti-mouse and anti-goat Cy3 secondary antibodies (1/100) were used to detect immunoreactive protein. The sections were imaged on a Zeiss Axiophot microscope interfaced with a SPOT camera (Diagnostic Instruments, Sterling Heights, Michigan) and a Gateway computer (N. Sioux City, SD).

Two-dimensional gels: Tissues were snap frozen in liquid nitrogen and ground to a fine powder using mortar and pestle. The proteins were solubilized in 100 mM DTT, 6 M Urea, 2% CHAPS overnight. 30 µg of protein was loaded onto 12 x 15 cm slab isofocusing gels consisting of 4% acrylamide, 0.1% piperazine diacrylamide, 9 M urea, 5 % ampholines (5 parts 6-8, 3 parts 5-7, and 2 parts 3-10), 2% CHAPS. The cathode buffer consisted of 20 mM sodium hydroxide and the anode buffer 10 mM

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phosphoric acid. The proteins were focused for 10,000 volt hours. The gels were fixed in 10% trichloroacetic acid and stained overnight with Neuhoff's Coomassie stain. The lanes of stained proteins were cut from the isofocusing slab gels and equilibrated in 10 mM Tris (pH 6. 8), 3% SDS, 19 % ethanol, 4% β-mercaptoethanol, and 0.004% bromophenol blue, for 10 minutes. The proteins were then separated on 12% acrylamide SDS gels and transferred to Immobi1 on 100 mAmp for 12 hours. The blots were probed for HSP20 as described above. The isoelectric focusing gradient was determined with BioRad IEF standards.

Peptide sequencing: The peptides, HSP20 phospho serine analogue N-WLRRASphosAPLPGLK (HSP20-PS) and a scrambled phosphorylated peptide N-PRKSphosLWALGRPLA (HSP20-SC) were synthesized on a Procise (Applied Biosystems, Model 492) instrument using standard protocols. The peptides were purified using high pressure liquid chromatography and purity was assured with mass spectrometry as previously described.

Permeabilization of Cardiac Myocyte: Permeabilization of cardiac myocytes was performed as described (11). Briefly, cells were slowly cooled by sequential two minute incubations with room temperature PBS then with 40°C PBS in an ice bath for two minutes. The PBS was discarded, and the cells were incubated with ATP (30 μ1 of 200mmol/L ATP, pH 7.4) followed immediately by permeabilization buffer (20 mmol/L HEPES, pH 7.4, 10 mmol/L EGTA, 140 mmol/L KCl, 50g/mL saponin, 5mmol/L oxalic acid dipotassium salt) containing the peptides (10μM) for 10 minutes in an ice bath. The cells were then washed four times on ice with chilled PBS. The cells were then returned to 37°C by incubations with room temperature PBS and 37°C PBS. The original cell media was then added at 37 °C.

25 Measurement of Cardiac Myocyte Contraction Rate/Relaxation Rate: The culture dishes containing the myocytes were placed on a Harvard Apparatus temperature regulation device positioned on the stage of an inverted microscope (Carl Zeiss Inc., Munchen- Hallbergmoos, Germany) and maintained at 37°C with a jacketed water bath. The microscope was outfitted with a digital camera and Video Savant software.
30 To determine the effects of HSP20 peptide analogues on contractile rate, individual cells were monitored before and after permeabilization. Contractile rates were

determined every 2 minutes (for a 15 second period) for a total of 10 minutes. Images of individual cells were captured at 30 frames/second.

Calcium Transients: Ca²⁺ transients were measured as previously described in detail elsewhere with only minor modifications (4). For these experiments myocytes were cultured on 25 mm glass coverslips. Following permeabilization (as described above), each coverslip was mounted in a leak-proof circular holding chamber (Medical Systems Inc., Cambridge MA) and gently washed several times with a Ringer solution containing: 142. mM NaCl, 4.0 mM KCl, 1.8 mM MgCl₂, 1.8 mM CaCl₂, 5.0 mM N-2-hydroxyethypiperazine-N-2-ethanesulfonic acid (HEPES, pH 7. 0), and 5.0 mM glucose. After washing, the myocytes were incubated in 1 ml Ringer solution with 1 µM fura-2 AM for 10 minutes at 37°C in a rotating water bath. The myocytes were subsequently washed several times with dye-free Ringer solution and allowed to stand covered at room temperature for 30 minutes to facilitate de-esterification.

Fluorescence measurements in individual rhythmically beating myocytes were carried out at 37°C using a DeltaScan microspectrofluorometer (Photon Technology International, NJ) coupled to an Olympus IX70 microscope equipped with an Olympus UApo/340 40X oil immersion objective with a numerical aperture of 1.35. The fura-2 transients reported are the ratio of Ca²⁺ fluorescence transients measured at excitation wavelengths 340 and 380 nm. The myocytes were only illuminated at short intervals, and each sample preparation was used for less than 1 hr to reduce the possibility of photobleaching or fura-2 leakage.

Statistical Analysis: Values are reported as mean +/- SEM, and "n" refers to the number animals examined. The statistical differences between two groups was determined with student's t test and between multiple groups with one way repeated measures analysis of variance (ANOVA) using Sigma Stat software (Jandel Scientific, San Rafeal, CA). A p-value less than 0.05 was considered significant.

Example 2

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This example illustrates the specificity of small HSP antibodies. The small heat shock proteins are highly homologous. Thus, to determine the specificity of the antibodies, immunoblots of recombinant HSP20, recombinant HSP25, bovine lens α B-crystallin, and homogenized rat heart were probed. Rat heart tissue homogenates (100 μ g), bovine α B-crystallin (1 μ g), recombinant HSP25 (1 μ g), and recombinant

HSP20 (1 μg) were separated by SDS-PAGE and transferred to Immobilion as described in the materials and methods. The blots were then probed with anti-HSP20 antibodies (1: 1000 dilution), anti-αB-crystallin antibodies (1: 1000 dilution) or anti-MKBP antibodies (1: 1000 dilution).

The HSP20, αB-crystallin, and MKBP antibodies recognized only the corresponding purified or recombinant protein. The anti-HSP20 antibodies recognized a major band at 20 kDa with a smaller band at a slightly lower relative mobility in heart homogenate proteins. This additional lower band may represent cross-reactivity with other proteins or may represent HSP20 that has been post translationally modified (eg. phosphorylated and nonphosphorylated HSP20). The anti-αB-crystallin antibodies recognized a single band with a relative mobility of 20 kDa in heart homogenates. The anti-MKBP antibodies recognized a band with a relative mobility of 20 kDa and another band with a higher relative mobility (approximately 35 kDa) in heart homogenates. The additional band may represent cross-reactivity with other proteins. There were very low levels of immunoreactive HSP25 in adult rat heart using an affinity purified mouse monoclonal antibody (20) or commercially available antibodies. These commercially available antibodies all recognized recombinant HSP25. Thus, only the associations between HSP20, αB-crystallin, and MKBP were examined in subsequent studies.

20 Example 3

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This example illustrates the subcellular localization and macromolecular associations of the small HSPs. To determine the intracellular distribution of the small heat shock proteins in cardiac myocytes, subcellular fractionation was performed as described in the materials and methods. Rat heart tissues were homogenized as described in the materials and methods. Blots were prepared with lanes containing proteins from the 10,000 x g supernatant, 10,000 x g pellet, 100,000 x g supernatant, and 100,000 x g pellet. The blots were probed with anti-HSP20 antibodies (1:1000 dilution), anti- α B-crystallin antibodies (1:1000 dilution), or anti- MKBP antibodies (1:1000 dilution).

The data from these experiments showed that HSP20 and MKBP were in the cytosolic fraction in rat heart homogenates. αB -crystallin was predominantly in the

cytosolic fraction but there was also a minor component of immunoreactive αB crystallin in the particulate fraction.

Since the small HSPs were predominantly cytosolic proteins, gel filtration was performed on the cytosolic fractions of rat heart muscle homogenates using a molecular sieving column (Superose 6). Fractions from the column were dot blotted with antibodies against HSP20, αB-crystallin, and MKBP. HSP20 and αB-crystallin were found in similar fractions that eluted from the column after the catalase standard (MW 232 kDa). MKBP eluted from the column in the same fraction as the bovine serum albumin standard (MW 67 kDa).

These data demonstrate that HSP20 and β -crystallin are associated in macromolecular aggregates in cardiac myocytes.

Example 4

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This example illustrates the cellular localization of the small HSP's using immunofluorescence microscopy Rat hearts were perfusion fixed and sections were stained with anti-sarcomeric actin antibodies (1:100 dilution), anti -αB-crystallin antibodies (1:100 dilution), anti-HSP20 antibodies (1:100 dilution), or anti-MKBP antibodies (1:100 dilution). Cy3 conjugated secondary antibodies were used and the slides were imaged with a fluorescence microscope (magnification, 63X).

Immunoreactive sarcomeric actin was present in distinct transverse bands. Immunoreactive αB -crystallin was also present in transverse bands but the staining was more punctate. Immunoreactive HSP20 was present in transverse bands and there was also staining of the cell membranes. Immunoreactive MKBP staining was less distinct, but there appeared to be longitudinal band-like staining with a distinct lack of immunoreactive staining for MKBP at the intercalated discs. Immunoreactive HSP25 was not detected using any of the 4 HSP25 antibodies tested. There was no specific pattern of immunoreactivity using pre-immune serum or no primary antibody.

These data demonstrate that crystalline and HSP20 have a staining pattern of distinct transverse bands similar to the pattern observed after staining for sarcomeric actin. This suggests that HSP20 is localized to the actin sarcomere.

30 Example 5

This example illustrates that activation of cyclic nucleotide signaling pathways leads to increases in the phosphorylation of HSP20 in cardiac myocytes. To determine

if activation of cyclic nucleotide signaling pathways led to increases in the phosphorylation of HSP20, two-dimensional immunoblotting was performed. Increases in the phosphorylation of HSP20 in vascular smooth muscle led to a shift of HSP20 from a basic to more acidic isoforms (2).

Rat cardiac myocytes were equilibrated in bicarbonate buffer for one hour. The myocytes were then treated with buffer alone or with the NO donor, sodium nitroprusside (10 μ M, 10 minutes). The cells were then homogenized, solubilized then separated by 2-dimensional electrophoresis. The proteins were then transferred to Immobilon then probed with anti-HSP20 antibodies (1:1000 dilution).

Blots from cells treated with buffer alone demonstrated a single point of immunoreactive protein with a relative mobility of 20 kDa and an isoelectric focusing point of 6.5. Blots from strips treated with sodium nitroprusside demonstrated a single point of immunoreactive protein with a relative mobility of 20 kDa and an isoelectric focusing point of 5.6. These isoelectric values are consistent with non-phosphorylated and phosphorylated HSP20 respectively.

Example 6

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This example illustrates that phosphorylated peptide analogues of HSP20 led to increase in the contractile rate of cardiac myocytes. To determine if HSP20 has a direct role in myocyte function, a model of transient permeabilization of cultured myocytes was used (11). The basal contractile rate of cardiac myocytes was determined using computer-assisted digital microscopy imaging techniques. The cells were then transiently permeabilized and incubated with N-PRKSphosLWALGRPLA (SEQ ID NO:299) (SC, a scrambled phosphorylated peptide) or with N-WLRRASphosAPLPGLK (PS, the phosphoHSP20 peptide analogue) (SEQ ID NO:300). Following permeabilization, the cells were placed back on the microscope stage and the contractile rate was again determined. The data represent the % increase in contraction rate over basal rate (%increase in rate, n=25 cells per group from 5 different experiments, * = P<0.05). Using a digital camera and Video Savant software, images of individual cells were captured at 30 frames/second before and after permeabilization and introduction of the PS peptide analogue.

Introduction of the phosphorylated peptide analogue of HSP20 (SEQ ID NO:300) resulted in a significant increase in the rate of cardiac myocyte contraction (37.6% +/- 2.4%). Similar increases in the rate of contraction occurred after treatment

with sodium nitroprusside (10 μ M, 33. 5% +/- 5.3%). In experiments where the myocytes were permeabilized in the absence of peptides (1. % +/- 0.5% increase) or scrambled phosphorylated peptides (5.9% +/- 1.9% increase) (SEQ ID NO:299), there was no significant change in the contractile rate.

Since phosphorylated HSP20 is associated with relaxation of vascular smooth muscle, we hypothesized that phosphopeptide analogues of HSP20 (SEQ ID NO:300) increased cardiac myocyte contractile rate by increasing their relaxation rate. To determine the rate of relaxation of the cardiac myocytes, contractile properties were analyzed using a digital camera and Video Savant software. The times for contraction and relaxation were measured before and after transient permeabilization in the presence of peptide analogues. There was a 50% increase in the time of relaxation in the cells exposed to the phosphopeptide analogue of HSP20 (SEQ ID NO:300). The relaxation time went from 0.06 seconds to 0.03 seconds. There were no significant changes in the relaxation rate in myocytes permeabilized in the absence of peptide, (0.058 +/- 0.005 s) or incubated in the presence of the scrambled phosphorylated peptide (0.055 +/- 0.006s).

Example 7

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We next examined Ca^{2+} fluxes in transiently permeabilized myocytes using the Ca^{2+} fluoroprobe, fura-2. Myocytes cultured on glass coverslips were permeabilized in the presence of a phosphopeptide analogue of HSP20 (WLRRASAPLPGLK with a phosphoserine, SEQ ID NO:300) or scrambled phosphopeptides (PRKSLWALGRPLA with a phosphoserine SEQ ID NO:299) or permeabilized in the absence of peptides just prior to loading with fura-2 AM. Data were collected from individual rhythmically beating myocytes (ratio of light emitted at 510 nm when alternately excited at 340 and 380 nm). For each myocyte, data were collected for 20 - 25 seconds and used only if the basal level and the peak magnitudes of the Ca^{2+} transients remained stable over this period. For each data set, the time constants for the decay (τ) were determined for 5 consecutive Ca^{2+} transients and averaged. The data were collected from 16 - 23 myocytes per group in 3 separate experiments (see text; * =P < 0.05).

The results demonstrated a significant decrease in the time constant for exponential (τ) decay of the Ca²⁺ transients in the phosphoserine group that was permeabilized with the phosphopeptide analogue of HSP20 (0.104 +/- 0.005 sec., n =

16) compared to the group treated with the scrambled phosphopeptide (0.236 +/-0.011 sec., n=18) or the myocytes permeabilized in the absence of any phosphopeptide (0.242 +/- 0.021 sec., n=23), when determined by fitting the declining phase of the transient by a first order exponential. There was not a significant difference when comparing the τ decay in myocytes treated with scrambled phosphopeptide to the myocytes permeabilized in the absence of peptide. The decreased τ decay of the Ca²⁺ transient in the presence of phosphopeptide HSP20 analogue was consistent with the observation described above that demonstrated an increased rate of contractile relaxation when measured with digital photography. Moreover, the more rapid decline of the Ca²⁺ transient suggests that the increased relaxation rate is due to more rapid uptake of Ca²⁺ by the sarcoplasmic reticulum (SR).

There were no differences in the peak systolic Ca²⁺ transients when myocytes that contained the HSP20 phosphopeptide analogue were compared to the scrambled phosphopeptide (P> 0.05). However, the permeabilization with either of the phosphopeptides resulted in a decrease in the peak magnitude of the Ca²⁺ transient that was evident when comparing myocytes permeabilized with scrambled phosphopeptide (1.05 +/- 0.02 340/380 ratio units) or phosphopeptide HSP20 analogue (1.01 +/- 02) to permeabilized myocytes in the absence of peptide (1.16 +/- 0.02; P <0. 05). These data indicated that permeabilization with phosphopeptides resulted in a non-specific decrease in the magnitude of the transients. The diastolic Ca²⁺ level was not significantly different in any of the three groups. From these data it can be concluded that phosphorylation of HSP20 does not specifically affect the magnitude of the Ca²⁺ transient.

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We claim:

1 1. A method for increasing the contractile rate in heart muscle comprising 2 administering to an individual in need thereof an amount effective to increase the contractile rate in heart muscle of one or more polypeptides comprising a sequence 3 4 according one or more of: 5 general formula I: (a) 6 $X1-X2-[X3-A(X4)APLP-X5-]_u-X6$ wherein X1 is absent or is one or more molecules comprising one or more 7 8 aromatic ring: 9 X2 is absent or comprises a transduction domain; X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1); 10 X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, 11 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs; 12 X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3, 13 wherein Z1 is selected from the group consisting of G and D; 14 Z2 is selected from the group consisting of L and K; and 15 Z3 is selected from the group consisting of S, T, and K; 16 X6 is absent or comprises a transduction domain; and 17 18 wherein u is 1-5; and 19 (b) general formula II: 20 J1-J2-[J3-A(J4)APLP-J5]_u-J6 wherein J1 is absent or is one or more molecules comprising one or more 21 22 aromatic ring; J2 is absent or comprises a cell transduction domain; 23 J3 is 0-14 amino acids of the sequence of heat shock protein 20 between 24 residues 1 and 14 of SEQ ID NO:298; 25 J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, 26 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs; 27 J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160 28 29 of SEQ ID NO:298; and 30 J6 is absent or comprises a cell transduction domain.

1 2. The method of claim 1 wherein increasing the contractile rate in heart muscle

- 2 comprises decreasing the heart muscle relaxation rate.
- 1 3. The method of claim 1 wherein the individual suffers from one or more of
- 2 bradyarrythmia, bradycardia, congestive heart failure, stunned myocardium,
- 3 pulmonary hypertension, and diastolic dysfunction.
- 1 4. The method of claim 1 wherein the one or more polypeptides comprise a
- 2 sequence according to general formula I.
- 1 5. The method of claim 1 wherein the one or more polypeptides comprise a
- 2 sequence according to general formula II.
- 1 6. The method of claim 1 wherein the one or more polypeptides consists of a
- 2 sequence according to general formula I.
- 1 7. The method of claim 1 wherein the one or more polypeptides consists of a
- 2 sequence according to general formula II.
- 1 8 The method of claim 4 wherein at least one or X2 and X6 comprises a
- 2 transduction domain.
- 1 9. The method of claim 5 wherein at least one or J2 and J6 comprises a
- 2 transduction domain.
- 1 10. A method for treating a heart muscle disorder comprising administering to an
- 2 individual suffering from one or more of bradyarrythmia, bradycardia, congestive
- 3 heart failure, stunned myocardium, pulmonary hypertension, and diastolic dysfunction
- 4 an amount effective to increase heart muscle contractile rate of one or more
- 5 polypeptides comprising a sequence according one or more of:
- 6 (a) general formula I:
- 7 $X1-X2-[X3-A(X4)APLP-X5-]_u-X6$
- wherein X1 is absent or is one or more molecules comprising one or more
- 9 aromatic ring;
- 10 X2 is absent or comprises a transduction domain;
- 11 X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);

12 X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,

- 13 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
- 14 X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,
- wherein Z1 is selected from the group consisting of G and D;
- 16 Z2 is selected from the group consisting of L and K; and
- 23 is selected from the group consisting of S, T, and K;
- 18 X6 is absent or comprises a transduction domain; and
- wherein u is 1-5; and
- 20 (b) general formula II:
- 21 $J1-J2-[J3-A(J4)APLP-J5]_u-J6$
- 22 wherein J1 is absent or is one or more molecules comprising one or more
- 23 aromatic ring;
- J2 is absent or comprises a cell transduction domain;
- J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
- 26 residues 1 and 14 of SEQ ID NO:298;
- 27 J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
- 28 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
- J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
- 30 of **SEQ ID NO:298**; and
- 31 J6 is absent or comprises a cell transduction domain.
 - 1 11. The method of claim 10 wherein the one or more polypeptides comprise a
- 2 sequence according to general formula I.
- 1 12. The method of claim 10 wherein the one or more polypeptides comprise a
- 2 sequence according to general formula II.
- 1 13. The method of claim 10 wherein the one or more polypeptides consists of a
- 2 sequence according to general formula I.
- 1 14. The method of claim 10 wherein the one or more polypeptides consists of a
- 2 sequence according to general formula II.
- 1 15. The method of claim 11 wherein at least one or X2 and X6 comprises a
- 2 transduction domain.

1 16. The method of claim 12 wherein at least one or J2 and J6 comprises a

- 2 transduction domain.
- 1 17. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from bradyarrythmia.
- 1 18. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from bradycardia.
- 1 19. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from congestive heart failure.
- 1 20. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from stunned myocardium.
- 1 21. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from pulmonary hypertension.
- 1 22. The method of any one of claims 10-16 wherein the individual is suffering
- 2 from diastolic dysfunction.
- 1 23. A method for preventing congestive heart failure, comprising administering to
- 2 an individual suffering from or who has had one or more of hypertension, anemia,
- 3 hyperthyroidism, aortic stenosis, aortic insufficiency, tricuspid insufficiency,
- 4 coarctation of the aorta, septal defects, pulmonary stenosis, tetralogy of Fallot;
- 5 arrythmias, myocardial infarction, cardiomyopathy, pulmonary hypertension, chronic
- 6 bronchitis, and emphysema an amount effective to prevent congestive heart failure of
- 7 one or more polypeptides comprising a sequence according one or more of:
- 8 (a) general formula I:
- 9 $X1-X2-[X3-A(X4)APLP-X5-]_u-X6$
- wherein X1 is absent or is one or more molecules comprising one or more
- 11 aromatic ring;
- 12 X2 is absent or comprises a transduction domain;
- 13 X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);
- 14 X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
- 15 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;

16	X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,
17	wherein Z1 is selected from the group consisting of G and D;
18	Z2 is selected from the group consisting of L and K; and
19	Z3 is selected from the group consisting of S, T, and K;
20	X6 is absent or comprises a transduction domain; and
21	wherein u is 1-5; and
00	(b) general formula II:
22	(,, 5
23	J1-J2-[J3-A(J4)APLP-J5] _u -J6
24	wherein J1 is absent or is one or more molecules comprising one or more
25	aromatic ring;
26	J2 is absent or comprises a cell transduction domain;
27	J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
28	residues 1 and 14 of SEQ ID NO:298;
29	J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
30	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
31	J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
32	of SEQ ID NO:298; and
33	J6 is absent or comprises a cell transduction domain.
1	24. A method for preventing pulmonary hypertension, comprising administering
2	to an individual suffering from one or more of chronic bronchitis, emphysema,
3	pulmonary embolism, and intestinal pulmonary fibrosis an amount effective to
4	prevent pulmonary hypertension of one or more polypeptides comprising a sequence
5	according one or more of:
6	(a) general formula I:
7	X1-X2-[X3-A(X4)APLP-X5-] _u -X6
8	wherein X1 is absent or is one or more molecules comprising one or more
9	aromatic ring;
10	X2 is absent or comprises a transduction domain;
11	X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);
12	X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
13	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;

14	X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,
15	wherein Z1 is selected from the group consisting of G and D;
16	Z2 is selected from the group consisting of L and K; and
17	Z3 is selected from the group consisting of S, T, and K;
18	X6 is absent or comprises a transduction domain; and
19	wherein u is 1-5; and
20	(b) general formula II:
21	J1-J2-[J3-A(J4)APLP-J5] _u -J6
	wherein J1 is absent or is one or more molecules comprising one or more
22	
23	aromatic ring; J2 is absent or comprises a cell transduction domain;
24	J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
25	
26	residues 1 and 14 of SEQ ID NO:298; J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
27	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
28	J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
29	
30	of SEQ ID NO:298; and
31	J6 is absent or comprises a cell transduction domain.
1	25. A method for preventing bradyarrythmia, comprising administering to an
2	individual suffering from one or more of coronary heart disease, and atheroma
3	formation, and/or that have had a myocardial infarction, an amount effective to
4	prevent bradyarrythmia of one or more polypeptides comprising a sequence according
5	one or more of:
6	(a) general formula I:
7	X1-X2-[X3-A(X4)APLP-X5-] _u -X6
8	wherein X1 is absent or is one or more molecules comprising one or more
9	aromatic ring;
10	X2 is absent or comprises a transduction domain;
11	X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);
12	X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
13	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;

14	X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,
15	wherein Z1 is selected from the group consisting of G and D;
16	Z2 is selected from the group consisting of L and K; and
17	Z3 is selected from the group consisting of S, T, and K;
18	X6 is absent or comprises a transduction domain; and
19	wherein u is 1-5; and
20	(b) general formula II:
21	J1-J2-[J3-A(J4)APLP-J5] _u -J6
22	wherein J1 is absent or is one or more molecules comprising one or more
23	aromatic ring;
24	J2 is absent or comprises a cell transduction domain;
25	J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
26	residues 1 and 14 of SEQ ID NO:298;
27	J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
28	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
29	J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
30	of SEQ ID NO:298; and
31	J6 is absent or comprises a cell transduction domain.
1	26. A method for preventing stunned myocardium, comprising administering to an
2	individual suffering from cardiac ischemia an amount effective to prevent stunned
3	myocardium of one or more polypeptides comprising a sequence according one or
4	more of:
5	(a) general formula I:
6	X1-X2-[X3-A(X4)APLP-X5-] _u -X6
7	wherein X1 is absent or is one or more molecules comprising one or more
8	aromatic ring;
9	X2 is absent or comprises a transduction domain;
10	X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);
11	X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
12	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
13	X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,
14	wherein Z1 is selected from the group consisting of G and D;

15	Z2 is selected from the group consisting of L and K; and
16	Z3 is selected from the group consisting of S, T, and K;
17	X6 is absent or comprises a transduction domain; and
18	wherein u is 1-5; and
19	(b) general formula II:
20	J1-J2-[J3-A(J4)APLP-J5] _u -J6
21	wherein J1 is absent or is one or more molecules comprising one or more
22	aromatic ring;
23	J2 is absent or comprises a cell transduction domain;
24	J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
25	residues 1 and 14 of SEQ ID NO:298;
26	J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
27	hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
28	J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
29	of SEQ ID NO:298; and
	J6 is absent or comprises a cell transduction domain.
30	00 10 abbent 01 002.p2.002 a com
30	27. A method for preventing diastolic dysfunction, comprising administering to an
	27. A method for preventing diastolic dysfunction, comprising administering to an
1	
1 2	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent
1 2 3	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according
1 2 3 4	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of:
1 2 3 4 5	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I:
1 2 3 4 5 6	 27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]_u-X6
1 2 3 4 5 6 7	 27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]_u-X6 wherein X1 is absent or is one or more molecules comprising one or more
1 2 3 4 5 6 7 8	 27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]_u-X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring;
1 2 3 4 5 6 7 8	 27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]_u-X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring; X2 is absent or comprises a transduction domain;
1 2 3 4 5 6 7 8 9	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]u-X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring; X2 is absent or comprises a transduction domain; X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1);
1 2 3 4 5 6 7 8 9 10	 27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]u-X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring; X2 is absent or comprises a transduction domain; X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1); X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
1 2 3 4 5 6 7 8 9 10 11	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-]u-X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring; X2 is absent or comprises a transduction domain; X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1); X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
1 2 3 4 5 6 7 8 9 10 11 12	27. A method for preventing diastolic dysfunction, comprising administering to an individual suffering from left ventricular hypertrophy an amount effective to prevent diastolic dysfunction of one or more polypeptides comprising a sequence according one or more of: (a) general formula I: X1-X2-[X3-A(X4)APLP-X5-] _u -X6 wherein X1 is absent or is one or more molecules comprising one or more aromatic ring; X2 is absent or comprises a transduction domain; X3 is 0, 1, 2, 3, or 4 amino acids of the sequence WLRR (SEQ ID NO:1); X4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine, hydroxyproline, phosphoserine analogs and phosphotyrosine analogs; X5 is 0, 1, 2, or 3 amino acids of a sequence of genus Z1-Z2-Z3,

- 17 X6 is absent or comprises a transduction domain; and
- wherein u is 1-5; and
- 19 (b) general formula II:
- 20 J1-J2-[J3-A(J4)APLP-J5]_u-J6
- 21 wherein J1 is absent or is one or more molecules comprising one or more
- 22 aromatic ring;
- J2 is absent or comprises a cell transduction domain;
- J3 is 0-14 amino acids of the sequence of heat shock protein 20 between
- 25 residues 1 and 14 of SEQ ID NO:298;
- J4 is selected from the group consisting of S, T, Y, D, E, hydroxylysine,
- 27 hydroxyproline, phosphoserine analogs and phosphotyrosine analogs;
- J5 is 0-140 amino acids of heat shock protein 20 between residues 21 and 160
- 29 of **SEQ ID NO:298**; and
- J6 is absent or comprises a cell transduction domain.
- 1 28. The method of any one of claims 23-27 wherein the one or more polypeptides
- 2 comprise a sequence according to general formula I.
- 1 29. The method of any one of claims 23-27 wherein the one or more polypeptides
- 2 comprise a sequence according to general formula II.
- 1 30. The method of any one of claims 23-27 wherein the one or more polypeptides
- 2 consists of a sequence according to general formula I.
- 1 31. The method of claim 23-27 wherein the one or more polypeptides consists of a
- 2 sequence according to general formula Π .
- 1 32. The method of claim 28 wherein at least one or X2 and X6 comprises a
- 2 transduction domain.
- 1 33. The method of claim 29 wherein at least one or J2 and J6 comprises a
- 2 transduction domain.

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03-227-PCT seq listing.ST25.txt SEQUENCE LISTING

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<210> 237
<211> 10
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<400> 237
Xaa Arg Arg Ala Tyr Ala Pro Leu Pro Gly 1 5 10
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<220>
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Xaa Arg Arg Ala Tyr Ala Pro Leu Pro Asp
1 5 10
<210> 239
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03-227-PCT seq listing.ST25.txt
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<400> 239
Xaa Arg Arg Ala Tyr Ala Pro Leu Pro Gly Leu
1 5 10
<210> 240
<211> 11
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03-227-PCT seq listing.ST25.txt
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<210>
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<210> 246
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Xaa Arg Arg Ala Tyr Ala Pro Leu Pro Asp Leu Ser
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<400> 250
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<210> 251
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Xaa Leu Arg Arg Ala Tyr Ala Pro Leu Pro Asp
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<211> 12
<212> PRT
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1 5 10
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1 10
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Xaa Leu Arg Arg Ala Tyr Ala Pro Leu Pro Asp Leu
<210> 256
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03-227-PCT seq listing.ST25.txt
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<400> 256
Xaa Leu Arg Arg Ala Tyr Ala Pro Leu Pro Asp Lys
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1 10
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<210> 259
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Xaa Leu Arg Arg Ala Tyr Ala Pro Leu Pro Gly Lys Ser
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03-227-PCT seq listing.ST25.txt
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<210> 263
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<210> 265
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<210> 266
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<220>
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Xaa is Phe, Tyr, or Trp
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03-227-PCT seq listing.ST25.txt
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<210> 267
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1 10
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<210> 269
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<400> 269
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1 10
<210> 270
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<220>
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<210> 271
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1 10
<210> 272
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<400> 272
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<220>
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03-227-PCT seq listing.ST25.txt
<223> Xaa is Phe, Tyr, or Trp
 <400> 273
Xaa Trp Leu Arg Arg Ala Tyr Ala Pro Leu Pro Gly Lys Ser
<210> 274
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<223> Description of artificial sequence: artificial peptide
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<221> MISC_FEATURE
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<223> Xaa is Phe, Tyr, or Trp
<400> 274
Xaa Trp Leu Arg Arg Ala Tyr Ala Pro Leu Pro Gly Lys Thr 10
<210> 275
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Xaa Trp Leu Arg Arg Ala Tyr Ala Pro Leu Pro Asp Leu Ser
<210> 276
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<220>
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<400> 276
Xaa Trp Leu Arg Arg Ala Tyr Ala Pro Leu Pro Asp Leu Thr
1 10
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03-227-PCT seq listing.ST25.txt
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03-227-PCT seq listing.ST25.txt <400> 280

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<210> 281 <211> 11 <212> PRT <213> Artificial

<220>

Description of artificial sequence: artificial peptide <223>

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Tyr Ala Arg Ala Ala Arg Gln Ala Arg Ala 1 10

<210> 282

<211> 34

<212> PRT <213> Artificial

<220>

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Glu Arg Pro Arg Ala Pro Ala Arg Ser Ala Ser Arg Pro Arg Arg Pro 20 25 30

Val Glu

<210> 283 <211> 27 <212> PRT <213> Artificial

<220>

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Gly Trp Thr Leu Asn Ser Ala Gly Tyr Leu Leu Gly Leu Ile Asn Leu 10 15

Lys Ala Leu Ala Leu Ala Lys Lys Ile Leu 20 25

<210> 284 <211> 12 <212> PRT

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<220>

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03-227-PCT seq listing.ST25.txt
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10 15
<210> 286
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<210> 287
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<212> PRT
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<212> PRT
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Ala Trp Ser Gln Pro
<210> 289
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03-227-PCT seq listing.ST25.txt

<212> PRT <213> Artificial

<223> Description of artificial sequence: artificial peptide

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Lys Ala Leu Ala Leu Ala Lys Lys Ile Leu 20 25

<210> 290 <211> 18 <212> PRT <213> Artificial

<223> Description of artificial sequence: artificial peptide

<400> 290

Lys Leu Ala Leu Lys Leu Ala Leu Lys Ala Ala Leu Lys 1 5 10 15

Leu Ala

<210> 291 <211> 21 <212> PRT <213> Artificial

<223> Description of artificial sequence: artificial peptide

<400> 291

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1 10 15

Lys Lys Arg Lys Val

<210> 292 <211> 16 <212> PRT <213> Artificial

<223> Description of artificial sequence: artificial peptide

<400> 292

Lys Ala Phe Ala Lys Leu Ala Ala Arg Leu Tyr Arg Lys Ala Gly Cys
1 10 15

<210> <211>

PCT/US2003/026366

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1 10 15 <210> 294 <211> 16 <212> PRT <213> Artificial <223> Description of artificial sequence: artificial peptide <400> 294 Ala Ala Phe Ala Lys Leu Ala Ala Arg Leu Tyr Arg Lys Ala Gly Cys $1 \hspace{1cm} 5 \hspace{1cm} 15$ <210> 295 <211> 16 <212> PRT <213> Artificial <220> <223> Description of artificial sequence: artificial peptide <400> Lys Ala Phe Ala Ala Leu Ala Ala Arg Leu Tyr Arg Lys Ala Gly Cys $1 \hspace{1cm} 5 \hspace{1cm} 10 \hspace{1cm} 15$ <210> 296 <211> 16 <212> PRT <213> Artificial <220> <223> Description of artificial sequence: artificial peptide <400> Lys Ala Phe Ala Lys Leu Ala Ala Gln Leu Tyr Arg Lys Ala Gly Cys $1 \hspace{1cm} 15$

<210> 297 <211> 15 <212> PRT <213> Artificial

Description of artificial sequence: artificial peptide

<400> 297

Gly Gly Gly Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg 1 5 10 15

03-227-PCT seq listing.ST25.txt

<210> 298 <211> 160

<212> PRT <213> Homo sapiens

<400> 298

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Ala Pro Leu Pro Gly Leu Ser Ala Pro Gly Arg Leu Phe Asp Gln Arg 20 25 30

Phe Gly Glu Gly Leu Leu Glu Ala Glu Leu Ala Ala Leu Cys Pro Thr $35 \hspace{1cm} 40 \hspace{1cm} 45$

Thr Leu Ala Pro Tyr Tyr Leu Arg Ala Pro Ser Val Ala Leu Pro Val 50 55 60

Ala Gln Val Pro Thr Asp Pro Gly His Phe Ser Val Leu Leu Asp Val 65 70 75 80

Lys His Phe Ser Pro Glu Glu Ile Ala Val Lys Val Val Gly Glu His 85 90 95

Val Glu Val His Ala Arg His Glu Glu Arg Pro Asp Glu His Gly Phe 100 105 110

Val Ala Arg Glu Phe His Arg Arg Tyr Arg Leu Pro Pro Gly Val Asp 115 120 125

Pro Ala Ala Val Thr Ser Ala Leu Ser Pro Glu Gly Val Leu Ser Ile 130 140

Gln Ala Ala Pro Ala Ser Ala Gln Ala Pro Pro Pro Ala Ala Ala Lys 145 150 155 160

<210> 299

<211> 13

<212> PRT

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<220>
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<220>

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<222> (4)..(4)
<223> Phosphorylated Ser

<400> 299

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<210> 300
<211> 13
<212> PRT
<213> Artificial
<220>
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<220>
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<222> (6)..(6)
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<400> 300

Trp Leu Arg Arg Ala Ser Ala Pro Leu Pro Gly Leu Lys $1 ext{0}$